

RESEARCH ARTICLE

# Enrichment and Broad Representation of Plant Biomass-Degrading Enzymes in the Specialized Hyphal Swellings of *Leucoagaricus gongylophorus*, the Fungal Symbiont of Leaf-Cutter Ants

Frank O. Aylward<sup>1,2aa\*</sup>, Lily Khadempour<sup>1,2</sup>, Daniel M. Tremmel<sup>1,2</sup>, Bradon R. McDonald<sup>1,2</sup>, Carrie D. Nicora<sup>3</sup>, Si Wu<sup>3ab</sup>, Ronald J. Moore<sup>3</sup>, Daniel J. Orton<sup>3</sup>, Matthew E. Monroe<sup>3</sup>, Paul D. Piehowski<sup>3</sup>, Samuel O. Purvine<sup>3</sup>, Richard D. Smith<sup>3</sup>, Mary S. Lipton<sup>3</sup>, Kristin E. Burnum-Johnson<sup>3</sup>, Cameron R. Currie<sup>1,2\*</sup>

**1** Department of Bacteriology, University of Wisconsin-Madison, Madison, Wisconsin, United States of America, **2** Great Lakes Bioenergy Research Center, University of Wisconsin, Madison, Wisconsin, United States of America, **3** Biological Sciences Division, Pacific Northwest National Laboratory, Richland, Washington, United States of America

<sup>aa</sup> Current address: Center for Microbial Oceanography Research and Education, University of Hawaii at Manoa, Honolulu, Hawaii, United States of America

<sup>ab</sup> Current address: Department of Chemistry and Biochemistry, University of Oklahoma, Norman, Oklahoma, United States of America

\* [faylward@hawaii.edu](mailto:faylward@hawaii.edu) (FOA); [currie@bact.wisc.edu](mailto:currie@bact.wisc.edu) (CRC)



**OPEN ACCESS**

**Citation:** Aylward FO, Khadempour L, Tremmel DM, McDonald BR, Nicora CD, Wu S, et al. (2015) Enrichment and Broad Representation of Plant Biomass-Degrading Enzymes in the Specialized Hyphal Swellings of *Leucoagaricus gongylophorus*, the Fungal Symbiont of Leaf-Cutter Ants. PLoS ONE 10(8): e0134752. doi:10.1371/journal.pone.0134752

**Editor:** Sean Brady, Smithsonian National Museum of Natural History, UNITED STATES

**Received:** March 12, 2015

**Accepted:** July 13, 2015

**Published:** August 28, 2015

**Copyright:** This is an open access article, free of all copyright, and may be freely reproduced, distributed, transmitted, modified, built upon, or otherwise used by anyone for any lawful purpose. The work is made available under the [Creative Commons CC0](https://creativecommons.org/licenses/by/4.0/) public domain dedication.

**Data Availability Statement:** Spectral counts for all proteins identified here are available in Dataset S1 here. The proteins discussed in this paper have been deposited as part of a previous publication online at the following url: [http://omics.pnl.gov/view/publication\\_1059.html](http://omics.pnl.gov/view/publication_1059.html).

**Funding:** Proteomics measurements were supported by the U.S. Department of Energy (DOE), Office of Biological and Environmental Research, Genomic Science Program under the Pacific Northwest National Laboratory (PNNL) Pan-omics Program, and

## Abstract

Leaf-cutter ants are prolific and conspicuous constituents of Neotropical ecosystems that derive energy from specialized fungus gardens they cultivate using prodigious amounts of foliar biomass. The basidiomycetous cultivar of the ants, *Leucoagaricus gongylophorus*, produces specialized hyphal swellings called gongyliidia that serve as the primary food source of ant colonies. Gongyliidia also contain plant biomass-degrading enzymes that become concentrated in ant digestive tracts and are deposited within fecal droplets onto fresh foliar material as ants incorporate it into the fungus garden. Although the enzymes concentrated by *L. gongylophorus* within gongyliidia are thought to be critical to the initial degradation of plant biomass, only a few enzymes present in these hyphal swellings have been identified. Here we use proteomic methods to identify proteins present in the gongyliidia of three *Atta cephalotes* colonies. Our results demonstrate that a diverse but consistent set of enzymes is present in gongyliidia, including numerous plant biomass-degrading enzymes likely involved in the degradation of polysaccharides, plant toxins, and proteins. Overall, gongyliidia contained over three quarters of all biomass-degrading enzymes identified in the *L. gongylophorus* genome, demonstrating that the majority of the enzymes produced by this fungus for biomass breakdown are ingested by the ants. We also identify a set of 40 of these enzymes enriched in gongyliidia compared to whole fungus garden samples, suggesting that certain enzymes may be particularly important in the initial degradation of foliar material. Our work sheds light on the complex interplay between leaf-cutter ants

were performed in the Environmental Molecular Science Laboratory, a U.S. DOE national scientific user facility at PNNL in Richland, WA. Battelle operates PNNL for the DOE under contract DE-AC05-76RLO01830. This work was also supported by National Science Foundation grants DEB-0747002, MCB-0702025, and MCB-0731822 to CRC and the DOE Great Lakes Bioenergy Research Center (DOE Office of Science BER DE-FC02-07ER64494).

**Competing Interests:** The authors have declared that no competing interests exist.

and their fungal symbiont that allows for the host insects to occupy an herbivorous niche by indirectly deriving energy from plant biomass.

## Introduction

Leaf-cutter ants of the genus *Atta* are prevalent consumers of foliar biomass in the New World tropics that play an important role in nutrient cycling and ecosystem engineering in many Neotropical habitats [1–3]. Rather than directly consuming the foliar material they collect, leaf-cutter ants use their forage to cultivate symbiotic gardens that they then consume for food [1,4]. Although the gardens cultivated by leaf-cutter ants contain a variety of microbes [5–10], the primary cultivar of these insects is the basidiomycetous fungus *Leucoagaricus gongylophorus*, which plays a major role in degrading the plant forage of the ants [1,11–15]. In addition to its biodegradative role, this fungus also produces the primary sustenance for leaf-cutter ants in the form of nutrient-rich hyphal swellings called gongylidia [1,16], thereby converting recalcitrant plant polymers into nutrients readily available to its host ants.

A central aspect of this insect-microbe symbiosis is the degradation of plant biomass, and a number of behaviors have evolved in these insects to help them both exclude unwanted microbial pests from their gardens and enhance the biomass degrading capacity of *L. gongylophorus* [17–20]. Two examples of these behaviors, termed weeding and fungus grooming, allow the ants to efficiently remove both patches of dead or contaminated fungus garden and selectively filter out spores of foreign fungi, respectively [17]. Moreover, entire fungus gardens are structured into different strata such that fresh foliar material is integrated only in top layers before being degraded in a step-wise process as it moved into lower layers [21,22]. When fresh foliar material is integrated into the top strata, the ants first macerate it into smaller pieces and deposit fecal droplets on top before inoculating fresh cultures of *L. gongylophorus* from older strata [1].

Previous work has shown that the fecal droplets of leaf-cutter ants contain plant biomass-degrading enzymes, which we define here to include Carbohydrate Active Enzymes (CAZymes), Fungal Oxidative Lignin-degrading Enzymes (FOLymes), and proteases [23–25]. It has been postulated in previous studies that the deposition of these droplets onto fresh foliar material serves as a form of “pretreatment” [26,27] that initiates the degradation process before *L. gongylophorus* is inoculated. Many of the plant biomass-degrading enzymes in ant fecal droplets have been shown to be derived from *L. gongylophorus* [28,29], and gene expression analyses have shown that some of these enzymes appear to be highly expressed in gongylidia [30,31], suggesting that the enzymes concentrated by *L. gongylophorus* in gongylidia are still active after passing through the digestive tract of the ants. Furthermore, analysis of the *Atta cephalotes* genome has shown a reduction in the total number of encoded proteases compared to other insects [32], suggesting that the loss of these proteases may have been an adaptation allowing *L. gongylophorus* enzymes to pass through their digestive systems and become concentrated intact in fecal droplets.

Although the enzymes concentrated by *L. gongylophorus* in gongylidia have been shown to have broad hydrolytic activity against a variety of plant polymers [23–25,31,33] and are likely of great import to plant biomass degradation in this insect-fungal symbiosis, the full complement of enzymes present in gongylidia is unknown. Here we used proteomic methods to identify proteins present in the gongylidia of three laboratory-reared *Atta cephalotes* colonies. Our results provide insight into the full spectrum of enzymes used by leaf-cutter ants to pretreat

foliar biomass and help clarify the extent of the co-evolutionary adaptations of leaf-cutter ants and their symbiotic fungus that underpin this ecologically-important symbiosis.

## Materials and Methods

### Sample collection

Samples of gongylidia were collected from three laboratory-reared colonies of *Atta cephalotes* fed a diet of maple (*Acer*) and oak (*Quercus*) leaves three times weekly. No field collections were performed in this study, and because all ant colonies were previously collected no specific permission was required for our experiments. Our analyses involved no endangered or protected species. Fungus garden material was taken from the middle strata of fungus gardens and gongylidia were collected using sterile forceps and a dissecting microscope set at 40X magnification. For each sample approximately 50 mg of gongylidia were placed into 50  $\mu$ L of sterile water in a 1.5  $\mu$ L microcentrifuge tube before the mixture was frozen at  $-80^{\circ}\text{C}$  prior to protein extraction and proteomic characterization.

### Preparation of gongylidia samples

To lyse cells, frozen gongylidia were placed in an ice cold mortar and pestle and ground under liquid nitrogen for several minutes before being transferred into centrifuge tubes with 0.1 mm zirconia beads and bead beat on a Bullet Blender (Next Advance, Averill Park, NY) for 3 minutes on speed 8. Samples were centrifuged and a methanol/chloroform extraction was performed to separate the protein, metabolites and lipids. Ice cold ( $-20^{\circ}\text{C}$ ) chloroform:methanol mix (prepared 2:1 (v/v)) was added to the sample in a 5:1 ratio over sample volume and vigorously vortexed. The samples were then placed on ice for 5 minutes and then vortexed for 10 seconds followed by centrifugation at  $10,000 \times g$  for 10 minutes at  $4^{\circ}\text{C}$ . The upper water soluble metabolite phase was collected into a glass vial, the lower lipid soluble phase was collected into a separate fresh glass vial, and both samples were dried in a SpeedVac before being stored at  $-80^{\circ}\text{C}$  until analysis. The remaining protein interlayer was placed in a fume hood to dry.

The protein pellets were resuspended in 100 mM  $\text{NH}_4\text{HCO}_3$  and assayed with Bicinchoninic acid (BCA) (Thermo Scientific, Rockford, IL) to determine the protein concentration. 2,2,2-Trifluoroethanol (TFE) (Sigma, St. Louis, MO) was added for a final concentration of 50% TFE. The samples were sonicated in an ice-water bath for 1 minute and incubated at  $60^{\circ}\text{C}$  for 2 hours with gentle shaking at 300 rpm. The samples were reduced with 2 mM Dithiothreitol (DTT) (Sigma, St. Louis, MO) with incubation at  $37^{\circ}\text{C}$  for 1 hour with gentle shaking at 300 rpm. Samples were then diluted 5-fold with 50 mM  $\text{NH}_4\text{HCO}_3$ , 1 mM  $\text{CaCl}_2$  for preparation for digestion. Sequencing-grade modified porcine trypsin (Promega, Madison, WI) was added to the sample at a 1:50 (w/w) trypsin-to-protein ratio for 3 hours at  $37^{\circ}\text{C}$ . The samples were concentrated in a SpeedVac (ThermoSavant, Holbrook, NY) to a volume of  $\sim 30 \mu\text{L}$ , acidified with trifluoroacetic acid to pH 4 and then centrifuged at  $10,000 \times g$ . The supernatant was cleaned with a strong cation exchange solid phase extraction procedure (SCX-SPE) on an OMIX tip (Agilent technologies, Santa Clara, CA) according to the manufacturer's instructions. The remaining pellet was re-digested, by resolubilization in 2M thiourea, 7 M urea and 1% CHAPS detergent and incubated at  $60^{\circ}\text{C}$  for 2 hours, reduced with 10mM Dithiothreitol (DTT) with incubation at  $37^{\circ}\text{C}$  for 1 hour, diluted 10-fold with 50 mM  $\text{NH}_4\text{HCO}_3$ , 1mM  $\text{CaCl}_2$ . The sample was then re-digested with trypsin for 3 hours at  $37^{\circ}\text{C}$  and cleaned with a strong cation exchange-solid phase extraction procedure (SCX-SPE). The supernatant and pellet samples were combined and concentrated down to  $\sim 30 \mu\text{L}$  using a SpeedVac and a final BCA assay was performed to determine the peptide concentration. The samples were then vialled for mass spectrometric analysis.

## Proteomic Analyses

Due to difficulties in collecting large quantities of gongylidia, on-line multidimensional fractions techniques were utilized in this study to increase measurement coverage given the small sample sizes available. Three biological replicates (5  $\mu\text{g}$  injections) of each sample were analyzed utilizing an on-line 2-D liquid chromatography (LC) separation consisting of one strong cation exchange (SCX) column and two reversed phase C18 columns and two solid phase extraction (SPE) columns for desalting. A total of 15 fractions were collected in this method, and while each fraction was collected, the previously collected fraction was analyzed by the analytical reversed phase separation with detection at the mass spectrometer. This system continuously acquired data for reversed phase separations through all 15 fractions. The 2D-LC system was custom built using two Agilent 1200 nanoflow pumps and one 1200 capillary pump (Agilent Technologies, Santa Clara, CA), various Valco valves (Valco Instruments Co., Houston, TX), and a PAL autosampler (Leap Technologies, Carrboro, NC). Full automation was made possible by custom software that allowed for parallel event coordination and therefore near 100% MS duty cycle through use of two trapping columns and two analytical columns. All columns were manufactured in-house by slurry packing media into fused silica (Polymicro Technologies Inc., Phoenix, AZ) using a 1-cm sol-gel frit for media retention [34]. A detailed schematic of this setup with setting configurations and reagent details is provided in Fig 1.

Mass spectrometry analyses were performed using a Velos Orbitrap mass spectrometer (Thermo Scientific, San Jose, CA) outfitted with a custom electrospray ionization (ESI) interface. Electrospray emitters were custom made by chemically etching 150  $\mu\text{m}$  o.d. x 20  $\mu\text{m}$  i.d. fused silica [35]. The heated capillary temperature and spray voltage were 275°C and 2.2 kV, respectively. Data were acquired for 100 minutes after a 10 minute delay from when the gradient started. Orbitrap spectra (AGC  $1 \times 10^6$ ) were collected from 400–2000  $m/z$  at a resolution of 100 k followed by data dependent ion trap CID MS/MS (collision energy 35%, isolation width 2.0, AGC  $1 \times 10^4$ ) of the ten most abundant ions. A dynamic exclusion time of 60 sec was used to discriminate against previously analyzed ions using a -0.6 to 1.6 Da mass window.

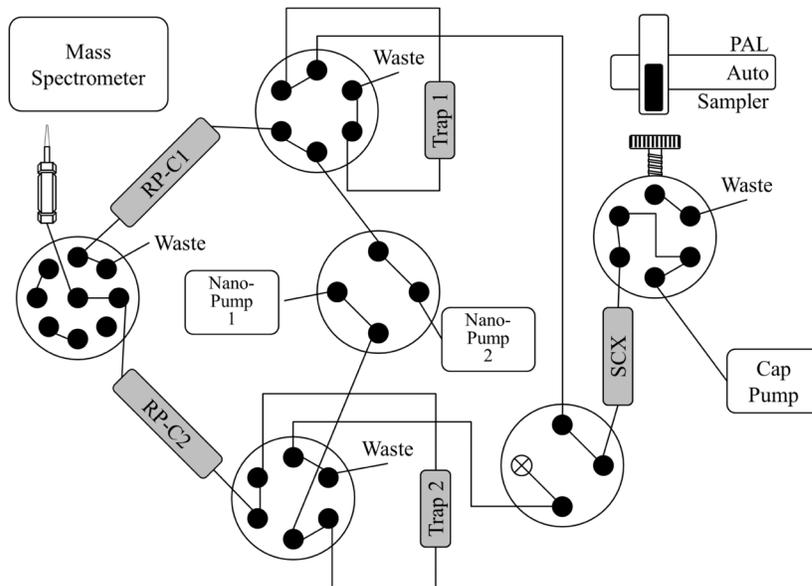
The peptide tandem mass spectra resulting from these procedures were mapped onto the *L. gongylophorus* genome [12] using MS-GF+ [36]. The resulting data were filtered by MS-GF  $< 1 \times 10^{-10}$  and mass error  $< \pm 2.5$  ppm which resulted in a spectral level FDR of 1.69% based on a decoy search (S1 Data). For each of the samples, peptide spectral counts were summed across the 15 fractions.

## Enzyme annotations and statistical analyses

Carbohydrate Active Enzymes (CAZymes, [37]), Fungal Oxidative Lignin-degrading enzymes (FOLymes, [38]), and protease annotations (following the MEROPs classification system [39]) for the *L. gongylophorus* genome were predicted and annotated to the protein family level, as previously described [12]. Additionally, proteins of interest in the gongylidia samples to which a large number of spectra could be mapped were annotated using homology searches against the Swissprot database [40] (downloaded 5/12/2014) using LAST version 393 [41] (minimum 70% length coverage required, top hits retained). Analysis of the *L. gongylophorus* proteins was performed using the gene predictions previously generated using GeneMark-ES [12,42].

Spectral counts for the *L. gongylophorus* proteins were compared to those previously reported from proteomic characterization of *Atta cephalotes* whole fungus gardens [12]. Only those spectra previously identified as mapping to *L. gongylophorus* proteins were used in this comparison. Fisher's Exact Test ( $P < 0.005$ ) was used to determine if the proportion of mass spectra mapping to plant biomass-degrading enzymes or their families was enriched in gongylidia compared to whole fungus gardens (top, middle, and bottom strata combined). For

Experimental Conditions						
Column Type	Ion Exchange		Trapping		Reversed Phase	
Dimensions	150 µm ID X 15 cm		150 µm ID X 5 cm		75 µm ID X 40 cm	
Packing Material	5 µm PolySULFOETHYLATM		5 µm Jupiter C18		3 µm Jupiter C18	
Mobile Phase	A: 0.1 mM NaH <sub>2</sub> PO <sub>4</sub> B: 3.0 mM NaH <sub>2</sub> PO <sub>4</sub>		A: 0.1% Formic Acid in H <sub>2</sub> O B: 0.1% Formic Acid in Acetonitrile			
Flow Rate	1.5 µL/min Capillary Pump		0.4 µL/min (Nano Pump)			
Gradients	Semi-Continuous Gradient				Linear Gradient	
	Fraction	%B	Fraction	%B	Time (min)	%B
	1	0.0-0.0	9	8.0-9.0	0	0
	2	0.0-1.5	10	9.0-10.0	2	10
	3	1.5-3.0	11	10.0-11.0	12	20
	4	3.0-4.0	12	11.0-14.0	85	35
	5	4.0-5.0	13	14.0-17.0	95	95
	6	5.0-6.0	14	17.0-20.0	100	95
	7	6.0-7.0	15	20.0-70.0	103	0
	8	7.0-8.0				



**Fig 1. Schematic of the 2D Liquid Chromatography Mass Spectrometry (2D-LC-MS) setup used in this study. SCX: Strong cation exchange.**

doi:10.1371/journal.pone.0134752.g001

ordination analyses raw spectral counts mapping to proteins identified in the gongyldia and fungus garden samples were normalized using the arcsine square-root transform. Non-metric multidimensional scaling (NMDS) was conducted on these transformed normalized counts using the vegan package in the R statistical programming environment [43,44].

## Results and Discussion

We identified a total of 636 *L. gongylophorus* proteins using 27,313 total spectral identifications (17,660 unique spectral identifications after quality-filtering [45]) in the three gongyldia

**Table 1. Summary of spectral counts and proteins identified.**

Gongylidia Sample	Total Mapped Spectra	Total Proteins Identified	CAZymes Identified	FOLymes Identified	Proteases Identified
Sample 1	2,531	388	32	24	29
Sample 2	9,007	275	30	19	27
Sample 3	15,775	498	37	28	38
Total	27,313	636	49	31	43

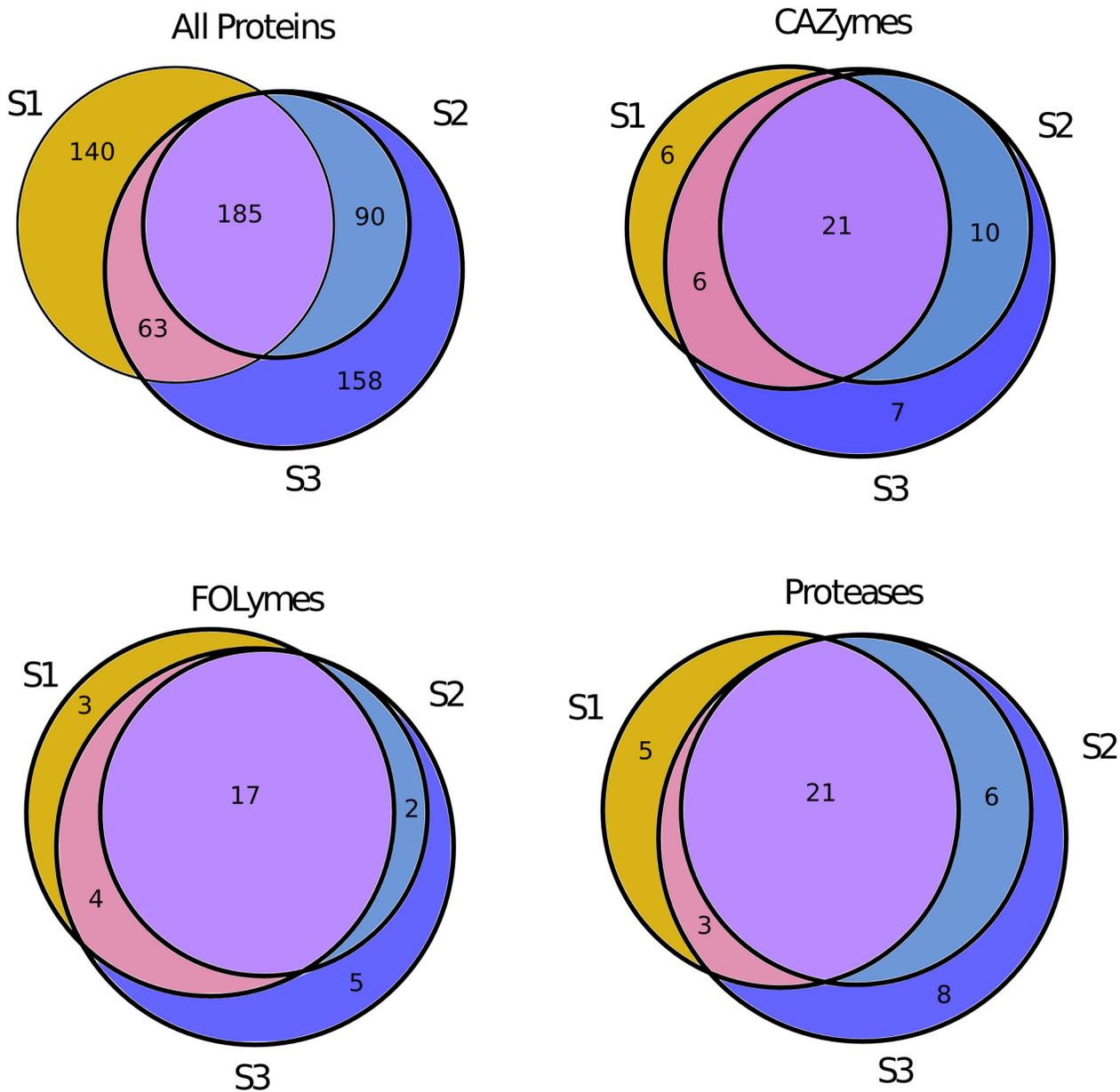
doi:10.1371/journal.pone.0134752.t001

samples tested (275–498 proteins identified per sample) (Table 1 and S1 Data). Of these proteins, 123 (19.3%) were predicted plant biomass-degrading enzymes including 50 Carbohydrate Active Enzymes (CAZymes) likely involved in polysaccharide degradation, 31 Fungal Oxidative Lignin-Degrading Enzymes (FOLymes) likely involved in aromatic compound degradation, and 43 proteases likely involved in protein degradation. One of these enzymes (LAG\_513) was found to have matches both to family GH5 glycoside hydrolases and family A22B aspartyl proteases. Of the 636 proteins identified, 185 (29%) were found in all three samples, while 338 (53%) were found in at least two samples (Fig 2). Many of the plant biomass-degrading enzymes identified were also shared between samples, with 21 CAZymes (42%), 17 FOLymes (55%), and 21 proteases (49%) identified in all three samples (Fig 2).

Spectra mapping to CAZymes, FOLymes, and proteases were over-represented in the gongylidia samples compared to spectra mapping to the same enzyme sets in earlier proteomic experiments performed on whole fungus garden material [12] (Fisher’s Exact Test,  $p < 0.005$ ). Moreover, nonmetric multidimensional scaling (NMDS) analyses of overall protein profiles of the gongylidia and whole fungus garden samples confirmed distinct clustering between these two sample groups (Fig 3). Together, these results indicate that the overall protein content of gongylidia is distinct from that of whole fungus gardens, and that plant biomass-degrading enzymes in particular are enriched in these hyphal swellings. The enrichment of plant biomass-degrading enzymes in gongylidia relative to whole fungus gardens, the large portion of shared proteins in the gongylidia samples (Fig 2), and the overall similarity of gongylidia protein profiles relative to those of whole fungus garden samples (Fig 3) all support the hypothesis that a consistent and specialized set of enzymes is produced in these hyphal swellings.

Comparisons of the spectral abundance of individual proteins between gongylidia and whole fungus gardens revealed that 129 proteins were enriched in at least one gongylidia sample, and 25 of these proteins were enriched in all three samples (Fisher’s Exact Test,  $p < 0.005$ ). The enzymes enriched in all three gongylidia samples include 6 CAZymes, 1 FOLyme, and 3 proteases, and another 13 CAZymes, 6 FOLymes, and 11 proteases were enriched in either one or two gongylidia samples (Fig 4). The 6 CAZymes enriched in all three gongylidia samples belong to enzyme families known to catalyze the degradation of pectin (family CE8) and glucosides such as starch (GH2, GH3, GH31). The three proteases enriched in all three gongylidia samples are metalloproteases (LAG\_100, LAG\_1037, and LAG\_1536), consistent with the identification of these enzymes in fecal droplets of *Atta texana* [23,28]. An LO1 laccase enriched in two gongylidia samples (LAG\_2404) has previously been implicated in the detoxification of plant defense compounds potentially harmful to *L. gongylophorus* or the host ants [31]. The aryl-alcohol oxidase (LAG\_4156) enriched in all three samples may play a role similar or auxiliary to laccases, although in other basidiomycetous fungi enzymes of this class have been implicated in hydrogen peroxide generation and lignocellulose degradation [46,47], and so it may serve to target structural polymers in foliar biomass rather than plant defense compounds.

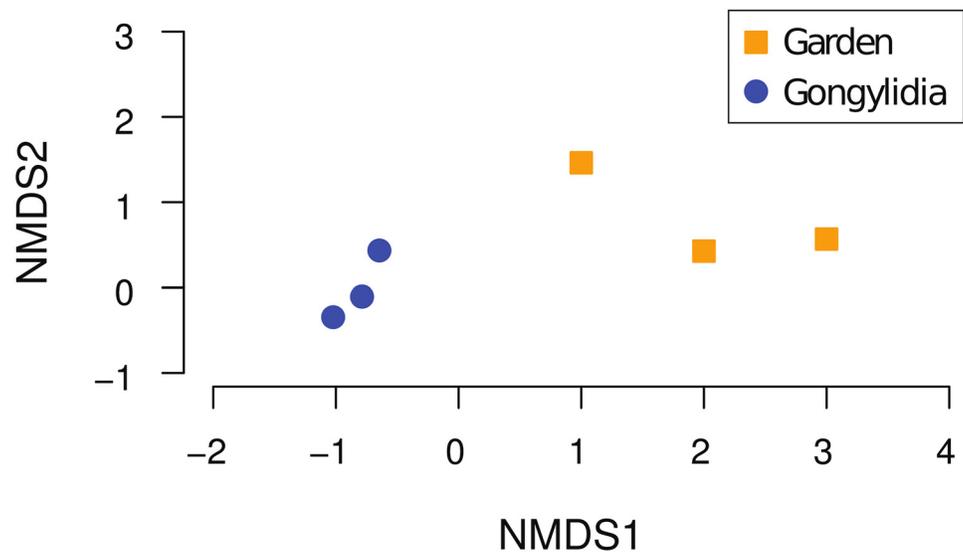
The specialized manner in which leaf-cutter ants cultivate fungi for food has been of great interest to scientists for decades [4], and a central goal of many studies has been to characterize



**Fig 2. Venn diagram showing shared total proteins, CAZymes, FOLymes, and proteases identified in the three gongyldia samples (labeled S1-S3) analyzed in this study.**

doi:10.1371/journal.pone.0134752.g002

the process through which plant biomass is degraded in fungus gardens and converted into usable nutrients for the host ants [7,12,21,22,48,49]. Many earlier studies of leaf-cutter ants of the genus *Atta* as well as other related fungus-growing ant genera revealed the presence of proteases, glycoside hydrolases, and laccases in the fecal droplets of the ants [23–26,28,33], and many of these studies noted the probable fungal origin of these enzymes. More recently, studies of the leaf-cutter ant *Acromyrmex echinatior* have identified pectinases in the fecal droplets and gongyldia [48], and transcripts encoding for laccases, glycoside hydrolases, and proteases have been identified in the gongyldia of this species [30,31]. Moreover, a great deal of other research has investigated plant biomass degradation in leaf-cutter ant fungus gardens more



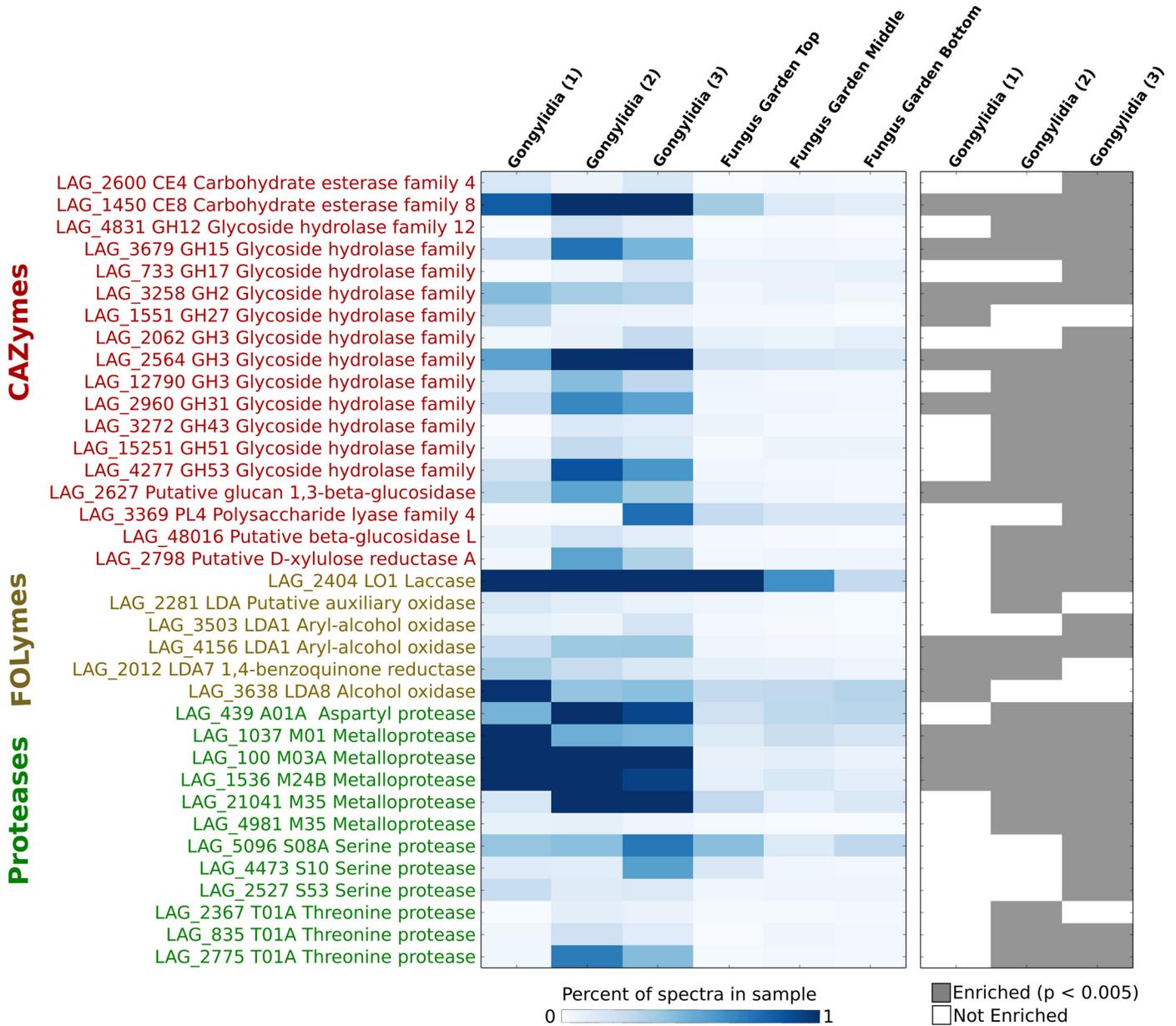
**Fig 3. Non-metric multidimensional scaling (NMDS) ordination plot of fungus garden and gongyldia samples.** Gongyldia samples are represented by blue circles, while orange squares represent fungus garden samples.

doi:10.1371/journal.pone.0134752.g003

generally without specific emphasis on gongyldia or fecal droplets [5,7,12,21,27,50–52]. Given the important role of gongyldia in supplying key biodegradative enzymes to the ants for subsequent deposition onto plant material, here we have built upon these previous studies by thoroughly identifying the CAZymes, FOLymes, and proteases present in the gongyldia of the leaf-cutter ant *Atta cephalotes* using novel proteomic methods.

Our finding of a broad array of plant biomass-degrading enzymes in the gongyldia of *L. gongylophorus* has important methodological implications for assaying the presence and activity of enzymes in leaf-cutter ant fungus gardens. Although a number of recent studies have relied on transcriptomics or mRNA quantification as measures of enzyme occurrence in fungus gardens [30,31,51], the enrichment of CAZymes, FOLymes, and proteases in gongyldia and the likely presence of a broad array of these enzymes in the fecal droplets deposited by the ants on fresh foliar biomass [23–25,28] indicates that transcription and protein presence are decoupled in this environment through the specialized gardening behavior of the leaf-cutter ants. This is likely common in fungus gardens since enzymes can be stored in gongyldia and transported away from where they are produced to sites where they are active. The measurement of mRNA alone is thus likely to provide incomplete and even misleading assessments of the presence of enzymes or their activities in specific locations within fungus gardens. We propose that for the inference of biodegradative processes it is more appropriate to use other techniques that rely on the direct measurement of proteins or their activities, such as the proteomic techniques used here and in other studies [12,48] or the direct measurements of enzymatic activity that have previously been used in fungus gardens [24,49,50].

Our finding that plant biomass-degrading enzymes are enriched in gongyldia supports the hypothesis that these enzymes play an important biodegradative role in leaf-cutter ant fungus gardens. Although only 10 CAZymes, FOLymes, and proteases were enriched in all three gongyldia samples, 36 plant biomass-degrading enzymes were enriched in at least one sample and 123 of these enzymes were identified in total. Recent analysis of the *L. gongylophorus* genome identified 145 putative plant biomass-degrading enzymes, and our finding of 118 of these enzymes (81% of the total, excluding the five glycoside hydrolases not reported previously)



**Fig 4. Heatmaps are presented that show the relative percent of total spectra that could be mapped to specific CAZymes, FOLymes, and proteases (left) and those enzymes that were found to be enriched in at least one gongyldia sample (right; Fisher's Exact Test,  $p < 0.005$ ). Only enzymes identified as enriched in at least one gongyldia sample compared to all fungus garden samples combined are shown.**

doi:10.1371/journal.pone.0134752.g004

provides the surprising result that the majority of the biodegradative enzymes produced by the fungal cultivar are ingested by the ants in some quantity. These results strongly implicate the ingestion of plant biomass-degrading enzymes and subsequent deposition on fresh foliar material within fecal droplets as a critical aspect of the degradation of plant biomass in fungus gardens. The initial pretreatment of biomass with a diverse set of enzymes prior to inoculation may release free sugars or more labile substrates critical for the initial establishment and sustained growth of *L. gongylophorus* in fungus gardens. The apparent lack of cellulases in gongyldia indicates that these enzymes are not part of the pretreatment step, possibly because

cellulose is used later in the degradation process. These results may partly explain why *L. gongylophorus* has been shown to grow slowly or not at all on numerous plant polymers in pure culture despite its apparent rapid growth in fungus gardens, where presumably the specialized gardening behaviors of the ants enhance biomass breakdown [1,11,14,53]. Together with these earlier results, our finding of widespread plant biomass-degrading enzymes in gongyldia therefore underscores the importance of the specialized gardening behaviors of leaf-cutter ants to biomass degradation as well as the extent to which leaf-cutter ants have co-evolved with their symbiotic fungus to efficiently derive energy from foliar material.

## Supporting Information

**S1 Data. Details regarding spectral identifications, the total number of spectra mapping to proteins in the samples, and enzyme annotations.**  
(XLSX)

## Acknowledgments

We thank members of the Currie lab for providing helpful comments on an earlier version of this manuscript and two anonymous reviewers for their helpful comments.

## Author Contributions

Conceived and designed the experiments: FOA LK KEB-J CRC. Performed the experiments: FOA DMT CDN SW RJM DJO PDP SOP. Analyzed the data: FOA LK BRM. Contributed reagents/materials/analysis tools: DMT CDN SW RJM DJO MEM PDP SOP RDS MSL. Wrote the paper: FOA LK KEB-J CRC.

## References

1. Weber NA. Fungus-Growing Ants. *Science*. 1966; 153: 587–604. doi: [10.1126/science.153.3736.587](https://doi.org/10.1126/science.153.3736.587) PMID: [17757227](https://pubmed.ncbi.nlm.nih.gov/17757227/)
2. Hölldobler B. The superorganism: the beauty, elegance, and strangeness of insect societies. 1st ed. New York: W.W. Norton; 2009.
3. Costa AN, Vasconcelos HL, Vieira-Neto EHM, Bruna EM. Do herbivores exert top-down effects in Neotropical savannas? Estimates of biomass consumption by leaf-cutter ants. *J Veg Sci*. 2008; 19: 849–854. doi: [10.3170/2008-8-18461](https://doi.org/10.3170/2008-8-18461)
4. Belt T. The naturalist in Nicaragua [Internet]. London, J.M. Dent & Sons, Ltd.; New York, E.P. Dutton & Co; 1911. Available: <http://archive.org/details/naturalistinnica01belt>.
5. Aylward FO, Burnum KE, Scott JJ, Suen G, Tringe SG, Adams SM, et al. Metagenomic and metaproteomic insights into bacterial communities in leaf-cutter ant fungus gardens. *ISME J*. 2012; 6: 1688–1701. doi: [10.1038/ismej.2012.10](https://doi.org/10.1038/ismej.2012.10) PMID: [22378535](https://pubmed.ncbi.nlm.nih.gov/22378535/)
6. Scott JJ, Budsberg KJ, Suen G, Wixon DL, Balsler TC, Currie CR. Microbial Community Structure of Leaf-Cutter Ant Fungus Gardens and Refuse Dumps. *PLOS ONE*. 2010; 5: e9922. doi: [10.1371/journal.pone.0009922](https://doi.org/10.1371/journal.pone.0009922) PMID: [20360970](https://pubmed.ncbi.nlm.nih.gov/20360970/)
7. Suen G, Scott JJ, Aylward FO, Adams SM, Tringe SG, Pinto-Tomás AA, et al. An Insect Herbivore Microbiome with High Plant Biomass-Degrading Capacity. *PLOS Genet*. 2010; 6: e1001129. doi: [10.1371/journal.pgen.1001129](https://doi.org/10.1371/journal.pgen.1001129) PMID: [20885794](https://pubmed.ncbi.nlm.nih.gov/20885794/)
8. Rodrigues A, Bacci M, Mueller UG, Ortiz A, Pagnocca FC. Microfungal “weeds” in the leafcutter ant symbiosis. *Microb Ecol*. 2008; 56: 604–614. doi: [10.1007/s00248-008-9380-0](https://doi.org/10.1007/s00248-008-9380-0) PMID: [18369523](https://pubmed.ncbi.nlm.nih.gov/18369523/)
9. Rodrigues A, Pagnocca FC, Bacci MJ, Hebling MJA, Bueno OC, Pfenning LH. Variability of non-mutualistic filamentous fungi associated with *Atta sexdens rubropilosa* nests. *Folia Microbiol (Praha)*. 2005; 50: 421–425.
10. Aylward FO, Suen G, Biedermann PHW, Adams AS, Scott JJ, Malfatti SA, et al. Convergent Bacterial Microbiotas in the Fungal Agricultural Systems of Insects. *mBio*. 2014; 5: e02077–14. doi: [10.1128/mBio.02077-14](https://doi.org/10.1128/mBio.02077-14) PMID: [25406380](https://pubmed.ncbi.nlm.nih.gov/25406380/)

11. Martin MM, Weber NA. The Cellulose-Utilizing Capability of the Fungus Cultured by the Attine Ant *Atta colombica tonsipes*. *Ann Entomol Soc Am*. 1969; 62: 1386–1387. PMID: [5374175](#)
12. Aylward FO, Burnum-Johnson KE, Tringe SG, Teiling C, Tremmel DM, Moeller JA, et al. *Leucoagaricus gongylophorus* Produces Diverse Enzymes for the Degradation of Recalcitrant Plant Polymers in Leaf-Cutter Ant Fungus Gardens. *Appl Environ Microbiol*. 2013; 79: 3770–3778. doi: [10.1128/AEM.03833-12](#) PMID: [23584789](#)
13. Chapela IH, Rehner SA, Schultz TR, Mueller UG. Evolutionary History of the Symbiosis Between Fungus-Growing Ants and Their Fungi. *Science*. 1994; 266: 1691–1694. doi: [10.1126/science.266.5191.1691](#) PMID: [17775630](#)
14. Siqueira CGD, Bacci M, Pagnocca FC, Bueno OC, Hebling MJ. Metabolism of Plant Polysaccharides by *Leucoagaricus gongylophorus*, the Symbiotic Fungus of the Leaf-Cutting Ant *Atta sexdens* L. *Appl Environ Microbiol*. 1998; 64: 4820–4822. PMID: [9835568](#)
15. Bacci M, Anversa MM, Pagnocca FC. Cellulose degradation by *Leucocoprinus gongylophorus*, the fungus cultured by the leaf-cutting ant *Atta sexdens rubropilosa*. *Antonie Van Leeuwenhoek*. 1995; 67: 385–386. doi: [10.1007/BF00872939](#) PMID: [7574556](#)
16. Bass M, Cherrett JM. Fungal hyphae as a source of nutrients for the leaf-cutting ant *Atta sexdens*. *Physiol Entomol*. 1995; 20: 1–6. doi: [10.1111/j.1365-3032.1995.tb00793.x](#)
17. Currie CR, Stuart AE. Weeding and grooming of pathogens in agriculture by ants. *Proc R Soc B Biol Sci*. 2001; 268: 1033–1039. doi: [10.1098/rspb.2001.1605](#)
18. Aylward FO, Currie CR, Suen G. The Evolutionary Innovation of Nutritional Symbioses in Leaf-Cutter Ants. *Insects*. 2012; 3: 41–61. doi: [10.3390/insects3010041](#)
19. Fernández-Marín H, Zimmerman JK, Nash DR, Boomsma JJ, Wcislo WT. Reduced biological control and enhanced chemical pest management in the evolution of fungus farming in ants. *Proc R Soc Lond B Biol Sci*. 2009; rspb.2009.0184. doi: [10.1098/rspb.2009.0184](#)
20. Fernández-Marín H, Zimmerman JK, Wcislo WT. Fungus garden platforms improve hygiene during nest establishment in *Acromyrmex* ants (Hymenoptera, Formicidae, Attini). *Insectes Sociaux*. 2007; 54: 64–69. doi: [10.1007/s00040-007-0907-z](#)
21. Kooij PW, Schiøtt M, Boomsma JJ, Licht HHDF. Rapid shifts in *Atta cephalotes* fungus-garden enzyme activity after a change in fungal substrate (Attini, Formicidae). *Insectes Sociaux*. 2011; 58: 145–151. doi: [10.1007/s00040-010-0127-9](#) PMID: [21475686](#)
22. Huang EL, Aylward FO, Kim Y-M, Webb-Robertson B-JM, Nicora CD, Hu Z, et al. The fungus gardens of leaf-cutter ants undergo a distinct physiological transition during biomass degradation. *Environ Microbiol Rep*. 2014; 6: 389–395. doi: [10.1111/1758-2229.12163](#) PMID: [24992538](#)
23. Boyd ND, Martin MM. Faecal proteinases of the fungus-growing ant, *Atta texana*: Properties, significance and possible origin. *Insect Biochem*. 1975; 5: 619–635. doi: [10.1016/0020-1790\(75\)90043-8](#)
24. Martin MM, Boyd ND, Gieselmann MJ, Silver RG. Activity of faecal fluid of a leaf-cutting ant toward plant cell wall polysaccharides. *J Insect Physiol*. 1975; 21: 1887–1892. doi: [10.1016/0022-1910\(75\)90219-X](#)
25. Martin MM, Gieselmann MJ, Martin JS. The presence of chitinase in the digestive fluids of ants. *Comp Biochem Physiol A Physiol*. 1976; 53: 331–332. doi: [10.1016/S0300-9629\(76\)80150-8](#)
26. Cherrett JM, Powell RJ, Stradling DJ. The Mutualism between leaf-cutter ants and their fungus. *Insect Fungal Interactions*. Waltham, MA: Academic Press; 1989. pp. 93–116.
27. Moller IE, De Fine Licht HH, Harholt J, Willats WGT, Boomsma JJ. The Dynamics of Plant Cell-Wall Polysaccharide Decomposition in Leaf-Cutting Ant Fungus Gardens. Chave J, editor. *PLOS ONE*. 2011; 6: e17506. doi: [10.1371/journal.pone.0017506](#) PMID: [21423735](#)
28. Boyd ND, Martin MM. Faecal proteinases of the fungus-growing ant, *Atta texana*: Their fungal origin and ecological significance. *J Insect Physiol*. 1975; 21: 1815–1820. doi: [10.1016/0022-1910\(75\)90247-4](#)
29. Rønne S, Boomsma JJ, Rosendahl S. Fungal enzymes transferred by leaf-cutting ants in their fungus gardens. *Mycol Res*. 2004; 108: 101–106. PMID: [15035511](#)
30. De Fine Licht HH, Boomsma JJ, Tunlid A. Symbiotic adaptations in the fungal cultivar of leaf-cutting ants. *Nat Commun*. 2014; 5: 5675. doi: [10.1038/ncomms6675](#) PMID: [25435021](#)
31. Licht HHDF, Schiøtt M, Rogowska-Wrzesinska A, Nygaard S, Roepstorff P, Boomsma JJ. Laccase detoxification mediates the nutritional alliance between leaf-cutting ants and fungus-garden symbionts. *Proc Natl Acad Sci*. 2013; 110: 583–587. doi: [10.1073/pnas.1212709110](#) PMID: [23267060](#)
32. Suen G, Teiling C, Li L, Holt C, Abouheif E, Bornberg-Bauer E, et al. The Genome Sequence of the Leaf-Cutter Ant *Atta cephalotes* Reveals Insights into Its Obligate Symbiotic Lifestyle. *PLOS Genet*. 2011; 7: e1002007. doi: [10.1371/journal.pgen.1002007](#) PMID: [21347285](#)

33. Rønshede S, Boomsma JJ, Rosendahl S. Fungal enzymes transferred by leaf-cutting ants in their fungus gardens. *Mycol Res.* 2004; 108: 101–106. doi: [10.1017/S0953756203008931](https://doi.org/10.1017/S0953756203008931) PMID: [15035511](https://pubmed.ncbi.nlm.nih.gov/15035511/)
34. Maiolica A, Borsotti D, Rappsilber J. Self-made frits for nanoscale columns in proteomics. *Proteomics.* 2005; 5: 3847–3850. doi: [10.1002/pmic.200402010](https://doi.org/10.1002/pmic.200402010) PMID: [16130174](https://pubmed.ncbi.nlm.nih.gov/16130174/)
35. Kelly RT, Page JS, Luo Q, Moore RJ, Orton DJ, Tang K, et al. Chemically etched open tubular and monolithic emitters for nano-electrospray ionization mass spectrometry. *Anal Chem.* 2006; 78: 7796–7801. doi: [10.1021/ac061133r](https://doi.org/10.1021/ac061133r) PMID: [17105173](https://pubmed.ncbi.nlm.nih.gov/17105173/)
36. Kim S, Gupta N, Pevzner PA. Spectral probabilities and generating functions of tandem mass spectra: a strike against decoy databases. *J Proteome Res.* 2008; 7: 3354–3363. doi: [10.1021/pr8001244](https://doi.org/10.1021/pr8001244) PMID: [18597511](https://pubmed.ncbi.nlm.nih.gov/18597511/)
37. Lombard V, Ramulu HG, Drula E, Coutinho PM, Henrissat B. The carbohydrate-active enzymes database (CAZy) in 2013. *Nucleic Acids Res.* 2014; 42: D490–D495. doi: [10.1093/nar/gkt1178](https://doi.org/10.1093/nar/gkt1178) PMID: [24270786](https://pubmed.ncbi.nlm.nih.gov/24270786/)
38. Levasseur A, Piumi F, Coutinho PM, Rancurel C, Asther M, Delattre M, et al. FOLy: an integrated database for the classification and functional annotation of fungal oxidoreductases potentially involved in the degradation of lignin and related aromatic compounds. *Fungal Genet Biol FG B.* 2008; 45: 638–645. doi: [10.1016/j.fgb.2008.01.004](https://doi.org/10.1016/j.fgb.2008.01.004) PMID: [18308593](https://pubmed.ncbi.nlm.nih.gov/18308593/)
39. Rawlings ND, Waller M, Barrett AJ, Bateman A. MEROPS: the database of proteolytic enzymes, their substrates and inhibitors. *Nucleic Acids Res.* 2014; 42: D503–D509. doi: [10.1093/nar/gkt953](https://doi.org/10.1093/nar/gkt953) PMID: [24157837](https://pubmed.ncbi.nlm.nih.gov/24157837/)
40. The UniProt Consortium. Activities at the Universal Protein Resource (UniProt). *Nucleic Acids Res.* 2014; 42: D191–D198. doi: [10.1093/nar/gkt1140](https://doi.org/10.1093/nar/gkt1140) PMID: [24253303](https://pubmed.ncbi.nlm.nih.gov/24253303/)
41. Frith MC, Hamada M, Horton P. Parameters for accurate genome alignment. *BMC Bioinformatics.* 2010; 11: 80. doi: [10.1186/1471-2105-11-80](https://doi.org/10.1186/1471-2105-11-80) PMID: [20144198](https://pubmed.ncbi.nlm.nih.gov/20144198/)
42. Borodovsky M, Mills R, Besemer J, Lomsadze A. Prokaryotic Gene Prediction Using GeneMark and GeneMark.hmm. *Current Protocols in Bioinformatics.* Hoboken, NJ: John Wiley & Sons, Inc.; 2002. Available: <http://onlinelibrary.wiley.com/doi/10.1002/0471250953.bi0405s01/abstract>.
43. R Core Team. R: A Language and Environment for Statistical Computing. Vienna, Austria; 2014.
44. Oksanen J, Blanchet FG, Kindt R, Legendre P, Minchin PR, O'Hara RB, et al. vegan: Community Ecology Package [Internet]. 2013. Available: <http://cran.r-project.org/web/packages/vegan/index.html>.
45. Piehowski PD, Petyuk VA, Sandoval JD, Burnum KE, Kiebel GR, Monroe ME, et al. STEPS: a grid search methodology for optimized peptide identification filtering of MS/MS database search results. *Proteomics.* 2013; 13: 766–770. doi: [10.1002/pmic.201200096](https://doi.org/10.1002/pmic.201200096) PMID: [23303698](https://pubmed.ncbi.nlm.nih.gov/23303698/)
46. Muheim A, Waldner R, Leisola MSA, Fiechter A. An extracellular aryl-alcohol oxidase from the white-rot fungus *Bjerkandera adusta*. *Enzyme Microb Technol.* 1990; 12: 204–209. doi: [10.1016/0141-0229\(90\)90039-S](https://doi.org/10.1016/0141-0229(90)90039-S)
47. Barrasa JM, Gutiérrez A, Escaso V, Guillén F, Martínez MJ, Martínez AT. Electron and Fluorescence Microscopy of Extracellular Glucan and Aryl-Alcohol Oxidase during Wheat-Straw Degradation by *Pleurotus eryngii*. *Appl Environ Microbiol.* 1998; 64: 325–332. PMID: [9435085](https://pubmed.ncbi.nlm.nih.gov/9435085/)
48. Schiøtt M, Rogowska-Wrzesinska A, Roepstorff P, Boomsma JJ. Leaf-cutting ant fungi produce cell wall degrading pectinase complexes reminiscent of phytopathogenic fungi. *BMC Biol.* 2010; 8: 156. doi: [10.1186/1741-7007-8-156](https://doi.org/10.1186/1741-7007-8-156) PMID: [21194476](https://pubmed.ncbi.nlm.nih.gov/21194476/)
49. Semenova TA, Hughes DP, Boomsma JJ, Schiøtt M. Evolutionary patterns of proteinase activity in attine ant fungus gardens. *BMC Microbiol.* 2011; 11: 15. doi: [10.1186/1471-2180-11-15](https://doi.org/10.1186/1471-2180-11-15) PMID: [21247468](https://pubmed.ncbi.nlm.nih.gov/21247468/)
50. De Fine Licht HH, Schiøtt M, Mueller UG, Boomsma JJ. Evolutionary Transitions in Enzyme Activity of Ant Fungus Gardens. *Evolution.* 2010; 64: 2055–2069. doi: [10.1111/j.1558-5646.2010.00948.x](https://doi.org/10.1111/j.1558-5646.2010.00948.x) PMID: [20067517](https://pubmed.ncbi.nlm.nih.gov/20067517/)
51. Grell MN, Linde T, Nygaard S, Nielsen KL, Boomsma JJ, Lange L. The fungal symbiont of *Acromyrmex* leaf-cutting ants expresses the full spectrum of genes to degrade cellulose and other plant cell wall polysaccharides. *BMC Genomics.* 2013; 14: 928. doi: [10.1186/1471-2164-14-928](https://doi.org/10.1186/1471-2164-14-928) PMID: [24373541](https://pubmed.ncbi.nlm.nih.gov/24373541/)
52. Lange L, Grell MN. The prominent role of fungi and fungal enzymes in the ant–fungus biomass conversion symbiosis. *Appl Microbiol Biotechnol.* 2014; 98: 4839–4851. doi: [10.1007/s00253-014-5708-5](https://doi.org/10.1007/s00253-014-5708-5) PMID: [24728757](https://pubmed.ncbi.nlm.nih.gov/24728757/)
53. Abril AB, Bucher EH. Evidence that the fungus cultured by leaf-cutting ants does not metabolize cellulose. *Ecol Lett.* 2002; 5: 325–328. doi: [10.1046/j.1461-0248.2002.00327.x](https://doi.org/10.1046/j.1461-0248.2002.00327.x)